

0040-4039(94)01664-X

Single Step Syntheses of Lactosylated Clusters by Telomerizations

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Abstract: N-Acryioylated lactoside derivatives containing spacer arms of different length were polymerized in the presence of thiols acting as radical scavengers to provide small oligomeric clusters (telomers) in a single step. t-Butyl mercaptan and methyl 3-mercaptopropionate, chosen as chain transfer reagents, were used for further attachment to L-lysine methyl ester.

Mammalian hepatic asialoglycoprotein receptors (ASGP-R) constitute cell surface lectins exhibiting high specificities toward non-reducing β -D-galactopyranosyl or 2-acetamido-2-deoxy- β -D-galactopyranosyl (Gal-GalNAc) residues present on glycoproteins and glycolipids.¹ These carbohydrate-protein interactions have been extensively studied in terms of carbohydrate specificities and requirements for multivalencies.² This last phenomenon, referred to as "glycoside cluster effect"³ is of paramount importance to achieve subnanomolar binding constants necessary for tight bindings. The routing of D-galactose-bearing clusters for the purpose of liver-specific drug delivery has been envisaged as viable anticancer and antiviral therapies.⁴ Furthermore, lactosylated clusters composed of galactoside residues were shown to be potent inhibitors of lung colonization by metastatic cancer cell lines.⁵

As previous lactosylated cluster syntheses were tedious and required multisteps, 3.69 we describe herein a direct, single step access to families of lactosylated clusters which depends on the polymerizations of Nacryloylated lactoside derivatives in the presence of thiols used as radical scavengers. Such process, called telomerizations, $\frac{10}{10}$ allows the chain propagation step to be quenched at an early stage to favor the formation of small repeating units (telomers). High molecular weight glycopolymers-bearing lactosides have been previously described. 11,12

Reduction of β -D-lactosyl azide 2, obtained in 98% yield under stereospecific phase transfer catalysis $(PTC)^{13.14}$ from α -acetobromolactose 1, afforded anomerically unstable lactosylamine 3 which was immediately treated with acryloyl chloride or with 6-N-acrylamidohexanoic acid and 2-ethoxy-1 ethoxycarbonyl-l,2-dihydroquinoline (EEDQ) in ethanol to provide protected monomer precursors 4 (85%) and 6 (73%) respectively. After Zemplén de-O-acetylation (NaOMe, MeOH) each resulting monomer 5 and 7 was telomerized in refluxing MeOH in the presence of 2,2'-azobisisobutyronitrile (AIBN) and *tert-butyl* mercaptan (t-BuSH) as first chain transfer reagent (telogen) in various molar ratios (Scheme 1). The reactions provided, in one single step, families of low molecular weight lactosylated telomers 8-9 which were readily separated by size exclusion chromatography on Bio-Gel P-2 column using water as eluent. In the case

of monomer 5, average number lactoside residues in the crude telomer mixtures (500 MHz $^{-1}$ H-NMR) ranged from 2-190 (by integration of the anomeric protons at 5.10 and 4.55 ppm relative to that of the t -butyl signal at 1.42 ppm) when using from 10-0.1 equivalents of t -BuSH. In a typical run with 10 equivalents of t -BuSH per monomer 5 (50 mg, 0.127 mmol) in 0.57 ml MeOH containing 0.2 mg AIBN (5 h, reflux), monoadduct 8 $(n=0, 29.4\%)$, dimer 8a $(n=1, 15.8\%)$, trimer 8b $(n=2, 8.4\%)$, tetramer 8c $(n=3, 5.8\%)$ and higher telomers 8d (n24, 41%) were obtained. Using 5.5 equivalents of t-BuSH per monomer 7, monoadduct 9 (26.6%), dimer 9a (13.9%), trimer 9b (1%) and higher telomers 9c-9d (45%) were also obtained in a total yield of 86.5%.

8 R' = t -Bu, $X = NH$, $n = 0$ (monomer), **8a** $n = 1$, **8b** $n = 2$, **8c** $n = 3$, **8d** n>3 9 R" = t-Bu, X = *NHCO(CIt2)sNH, n = O,* 9a n = 1,9b n = 2, 9c n = 3, 9d n>3 10 R' = *CH2CH2CO2Me, X = NHCCKCH2)sNH, n = O,* 10a n = 1,10b n = 2, 10¢ n =3, 10d n >3

Scheme 1. a: PTC, NaN₃, TBAHS, ETOAc, 1M Na₂CO₃, 30 min, 98%; b: H₂, Pd-C, EtOAc, o.n., 94%; c: CH₂=CHCOCl, pyridine, EtOAc, 45 min, 85%; d: HO₂C(CH₂)₅-NH-CO-CH=CHz, EEDQ, EtOH, r.t., 16 h, 73%; e: NaOMe, MeOH, r.t., 3 h, 98% (5) and 91% (7); f: t-BuSH (or HSCH₂CH₂ CO₂Me), AIBN, MeOH, reflux, 5h.

It is also worthy of mention that noticeable level of diastereoselectivities (ca. 2:1, as seen from the $¹$ H-</sup> NMR of 8a) were observed during the telomerizations of 5. This was not surprising because the monomer 5, which is deprived of spacer arm, was actually serving as chiral auxiliary to provide telomer tacticities.¹⁵

Similarly, when monomer 7 was telomerized with 0.43 equivalents of methyl 3-mercaptopropionate, monoadduct 10 (26%), dimer 10a (22%), trimer 10b (12.3%), tetramer 10c (7%) and higher telomers 10d $(n \geq 4, 30.8\%)$ were obtained.

To further exploit the usefulness of lactosylated telomers for drug attachment and dendrimer syntheses,¹⁶ model experiments with modified monomers 10 and 11 were examined independently. Peracetylated monomer 11, was obtained as a single product $(84%)$ when 6 was treated with a fifty fold molar excess of t-BuSH and a catalytic amount of AIBN in refluxing MeOH. Attempts to prepare 11 by nucleophilic attack of t-BuSH in the presence of triethylamine (Michael addition) failed. Successful deprotection of t-butyl thioether in 11 was accomplished in a two step process with 2-nitrobenzenesulfenyl chloride $(2-NO_2-PhSCI)^{17}$ in acetic acid to afford S-(2-nitrophenyl)sulfenyl derivative 12 in 84% yield. Disulfide reduction with 2-mercaptoethanol gave 13 (60%). The thiol 13 is suitably functionalized and can be covalently attached to N-chloroacetylated poly-L-lysine dendrimers.¹⁶ Treatment of methyl ester 10 with 0.05M NaOH provided acid 14 (quant). Preparation of a family of telomers related to 14 directly from 6 and 3-mercaptopropionic acid were hampered by difficulties in separation. Treatment of 14 (2 equiv) with lysine methyl ester and EEDQ as above afforded divalent lactosylated cluster 15 (50%) and its monomeric adduct (15%). Works are now in progress to conjugate each individual telomer with dendritic poly-L-lysine¹⁶ and other polyamines to provide rapid access to complex multivalent "dendritic telomers."

Scheme 2: a: 2-NO₂-Ph-SCl, HOAc, r.t., 3h, 84%; b: HSCH₂CH₂OH, MeOH, r.t., 3h, 60%; c: 0.05M NaOH, r.t., 3h, quant; d: 13, LysOMe, EEDQ, EtOH, DMF, 50%

Preliminary inhibition experiments with peanut lectin from *Arachis hypogaea* and telomers 8 deprived of spacer showed no improved binding properties as compared to those of lactose.¹⁸ On the other hand, trimer 10b was twice as potent as lactose.

Other model studies with galactosides containing N-acrylamidohexanoic acid as spacer arm provided analogs similar to 9. Using thiolacetic acid as radical scavenger gave complex telomer mixtures.

Acknowledgments: We thank the Natural Science and Engineering Research Council of Canada (NSERC) for financial support. We are also thankful to Dr. A. Romanowska from our laboratory for preliminary lectin assays.

References and Notes

- 1. Ashwell, G.; Harford, J. *Annu. Rev. Biochem.* 1982, *51,* 531-554.
- *2. Lee, Y. C. FASEB J.* 1992, 6, 3193-3200.
- 3. Lee, R. T.; Lee, Y. C. Enhanced Biochemical Affinities of Multivalent Neoglycoconiugates. In *Neoglycoconjugates: Preparation and Applications; Lee, Y. C.; Lee, R. T. Eds.; Academic Press: San* Diego, 1994; pp. 23-50.
- 4. Wu, G. Y.; Wu, C. H. In Liver Diseases: *Targeted Diagnosis and Therapy Using Specific Receptors and Ligands;* Wu, G. Y.; Wu, C. H. Eds.; Marcel Dekker: New York, 1991; pp. 127-149.
- 5. Dean, B.; Oguchi, H.; Cai, S.; Otsuji, E.; Tashiro, K.; Hakomori, S.-i.; Toyokuni, T. *Carbohydr. Res.* 1993, *245,* 175-192.
- 6. Lee., Y. C.; Lee, R. T.; Rice, K.; Ichikawa, Y.; Wong, T.-C. *Pure Appl. Chem.* 1991, *63,499-506.*
- 7. Haensler, J.; Schuber, F. *Glycoconjugate J.* 1991, 8, 116-124.
- 8. Krebs, A.; Depew, W. T.; Szarek, W. A.; Hay, G. W.; Hronowski, L. J. J. *Carbohydr. Res.* 1994, *254,* 257-268.
- 9. Findeis, M. A. *Int. J. Peptide Protein Res.* 1994, *43,477-485.*
- 10. Starks, C. M. *Free radical telomerization;* Academic Press: New York, 1974; pp. 1-3.
- 11. Roy, R.; Tropper, F. D.; Romanowska, A. *Bioconjugate Chem.* 1992, 3, 256-261.
- 12. Kobayashi, A.; Kobayshi, K.; Akaike, T. J. *Biomater. Sci. Polym. Educ.* 1992, 6, 499-508.
- 13. Tropper, F. D.; Andersson, F. O.; Braun, S.; Roy, R. *Synthesis* 1992, 618-620.
- 14. All new low molecular weight compounds gave satisfactory spectroscopic and analytical data.
- 15. Porter, N. A.; Swann, E.; Nally, J.; McPhail, *A. T. J. Am. Chem. Soc.* 1990, *112,* 6740-6741.
- 16 Roy, R.; Zanini, D.; Meunier, S. J.; Romanowska, A. J. *Chem. Sot., Chem. Commun.* 1993, 1869-1872.
- 17. Pastuszak, J. J.; Chimiak, A. J. Org. *Chem.* 1981, *46,* 1868-1873.
- 18. Park, W. K. C.; Aravind, S.; Romanowska, A.; Renaud, J.; Roy, R. *Methods Enzymol.* 1994, *242,* 292- 302.

(Received in USA 11 July 1994; *revised* 23 *August* 1994; *accepted* 26 *August* 1994)